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## HYDROGEN PEROXIDE CONCENTRATION AS AN INDICATOR OF ABIOTIC ENVIRONMENTAL STRESS OF MACROPHYTES FOR MANAGEMENT

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**Abstract.** Invasive species, *E. densa*, and Japanese native species, *Myriophyllum spicatum* and *Ceratophyllum demersum* were investigated in experimental tanks with different environmental conditions. Tissue hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) concentrations were measured responding to either individual or multiple environmental stressors of light intensity (PAR), and water temperature. The H<sub>2</sub>O<sub>2</sub> concentration increased in parallel to the increment of unpreferable levels of each abiotic factor, and the trend was independent of other factors. The total H<sub>2</sub>O<sub>2</sub> concentration is provided by the sum of contribution of each factor. Under increased total H<sub>2</sub>O<sub>2</sub> concentration chlorophyll-a (Chl-a) concentration of plant tissues decreases, then reduce their growth rate, and subsequently reduce their biomass. The H<sub>2</sub>O<sub>2</sub> concentration threshold, beyond which degradation is initiated, was between 16 and 20 μmol/gFW regardless of the type of environmental stresses. These results highlight the potential efficacy of total H<sub>2</sub>O<sub>2</sub> concentration as a proxy for the overall environmental condition. In Japanese waters, major environmental factors limiting macrophyte colonization were identified as water temperature, and high solar radiation. The relationship between the unpreferable levels of these factors and H<sub>2</sub>O<sub>2</sub> concentration was empirically obtained for these species. Then a mathematical model was developed to predict the colonization area of these species with environmental conditions. The tissue H<sub>2</sub>O<sub>2</sub> concentration decreases with increasing temperature for *E. densa* while increases for other species. Therefore, native species grow intensively in spring; however, they often deteriorate in summer. For *E. densa*, on the other hand, H<sub>2</sub>O<sub>2</sub> concentration decreases with high water temperature in summer, allowing intensive growth. High solar radiation, however, increases the H<sub>2</sub>O<sub>2</sub> concentration, deteriorating the plant in shallow zones.

**Keywords:** submerged macrophytes, environmental stress, hydrogen peroxide, stress indicator, invasive macrophytes

## 1. INTRODUCTION

Submerged macrophyte response to environmental conditions is species specific, and invasive plants tend to exhibit more tolerance than native species (Bates et al., 2013). Therefore, invasive species are able to dominate or distribute in areas where native species fail to survive. Among different invasive aquatic macrophytes, *Egeria densa* is a well-known worldwide species that causes significant ecological issues in freshwater ecosystems. In Japan, *E. densa* was introduced as an ornamental aquarium plant in the early 19th century, however, it had escaped into natural freshwater bodies and became naturalized in the 1940s, then, was mainly invaded into lakes in the last two decades. Japanese lakes were nearly free of macrophytes in 1960s, however, they are now intensively covered, though the mechanisms are unknown.

The investigation of submerged macrophyte behaviors is normally conducted by a long term monitoring. However, it is a hard work and costly (Biggs, et al, 2019; Vettori and Rice, 2020).

Plants suffer from oxidative stress when exposed to harsh environments, which results in an enormous amount of reactive oxygen species (ROS) in cells (Sharma et al. 2012). Under normal conditions, the activities of reactive oxygen species are balanced with antioxidants generated as a defense mechanism. However, under stressful conditions, antioxidant defenses become insufficient, resulting in oxidative stress that often leads to cell death (Choudhuri et al. 2017). Among the different types of ROS, superoxide anion ( $O_2^-$ ) and hydroxyl radicals ( $OH^-$ ) are the dominant species generated under stress and are routinely converted into relatively stable hydrogen peroxide ( $H_2O_2$ ), via either superoxide dismutase (SOD) or nonenzymatic systems (Asada 2006; Das et al. 2014). Thus, the concentration of  $H_2O_2$  has a strong potential to provide information regarding the environmental stress imposed on plants (Asaeda et al. 2018; Asaeda et al. 2020).

The objective of the current study is to understand the species specific preferable condition for colonization along the riparian zone by quantifying environmental stress using hydrogen peroxide concentrations.

## 2. MATERIALS AND METHODS

### 2.1 EXPERIMENTAL PROCEDURE

An experiment was conducted to identify the increment of  $H_2O_2$  concentration of the plant tissue under different water temperatures and irradiance levels (PAR), and, thereby, to make empirical relations between these factors. Several light levels (0–1,300  $\mu\text{mol}/\text{m}^2/\text{s}$  of PAR) were tested in small aquaria (dimensions: 50.0 cm  $\times$  35.0 cm  $\times$  35.0 cm). Temperature level was maintained at  $10 \pm 2$  (*E. densa*),  $15 \pm 2$  (*E. densa*),  $20 \pm 2$ ,  $25 \pm 2$  (*E. densa*), and  $30 \pm 2$ ,  $35 \pm 2$  (*E. densa*) $^\circ\text{C}$  using a temperature controlling system (Aquarium cooler ZC-100 $\alpha$ , Zensui Corporation, Tokyo, Japan). PAR intensity was irradiated under natural solar radiation or using LED lights (Model LT-NLD85L-HN, OHM Electric Inc., Japan) with a 12 h light:12 h dark photoperiod for 3 weeks. The length of *E. densa* grown in the experimental units was measured using a millimeter scale at 5–7 day intervals. The shoot growth rate (SGR) was calculated as the difference in shoot length between two observations divided by the duration, and it was expressed in cm/day.

### 2.2 ANALYSIS

Fresh plant shoots were extracted (~500 mg) in an ice-cold phosphate buffer (50 mM, pH 6.0) that

contained polyvinylpyrrolidone (PVP), and the extractions were centrifuged at  $5,000 \times g$  for 20 min at  $4^{\circ}\text{C}$ . This extraction was used to analyze the  $\text{H}_2\text{O}_2$  content spectrophotometrically following the  $\text{TiSO}_4$  method (Satterfield and Bonnell, 1955) with modifications. The reaction mixture contained 750  $\mu\text{l}$  of enzyme extract and 2.5 ml of 1%  $\text{TiSO}_4$  in 20%  $\text{H}_2\text{SO}_4$  (v/v), which was centrifuged at  $5,000 \times g$  for 15 min at  $20^{\circ}\text{C}$ . The optical absorption of the developed yellow color was measured spectrophotometrically at a wavelength of 410 nm. The  $\text{H}_2\text{O}_2$  concentration in samples was determined using the prepared standard curve for known concentration series and was expressed in  $\mu\text{mol}$  per gram fresh weight ( $\mu\text{mol/gFW}$ ). The results were compared with those of the e-FOX method, and suitable agreement was obtained (Queval et al., 2008).

Chlorophyll concentrations of experimental plants were determined spectrophotometrically (UV Mini 1210, Shimadzu, Japan) by extracting pigments with N,N-dimethylformamide after keeping them in darkness for 24 h, and they were expressed in terms of fresh weight (FW) (Wellburn, 1994; DeSilva and Asaeda 2017).

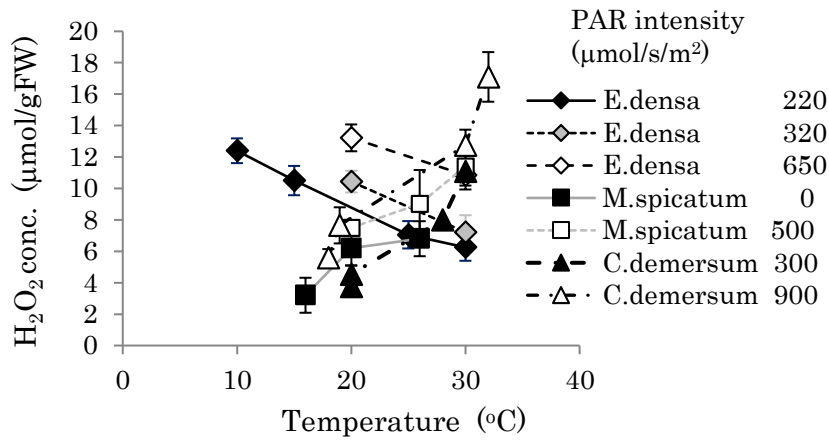
Data were tested for normality with the Shapiro–Wilk test before statistical analyses. All results were presented as the mean  $\pm$  SD of more than three replicates. Data were subjected to a one-way analysis of variance (ANOVA) with Tukey's post-hoc test for mean separation. The t-test was performed where necessary. Bivariate analysis was used and followed by Pearson's correlation to evaluate the relationship among parameters. Statistical analyses were performed in IBM SPSS V25.

### 3. RESULTS

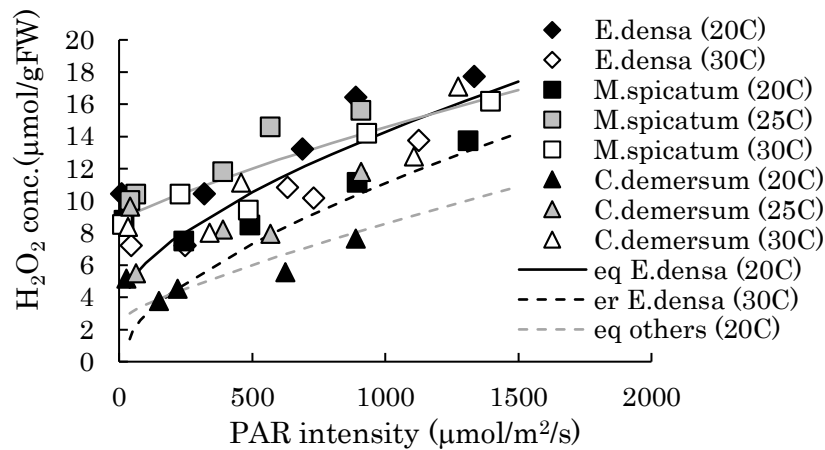
#### 3.1 Empirical relationships of $\text{H}_2\text{O}_2$ concentration with respect to abiotic factors

A species-specific response is shown on  $\text{H}_2\text{O}_2$  formation in macrophyte tissues with the combined stresses of temperature and light intensity (Figure 1). The basal  $\text{H}_2\text{O}_2$  concentrations were  $4.6 \mu\text{mol/gFW}$  at  $20^{\circ}\text{C}$  for *E. densa*, and  $3.0 \mu\text{mol/gFW}$  at  $20^{\circ}\text{C}$  for other species, respectively, after being exposed to dark conditions. The increment of  $\text{H}_2\text{O}_2$  driven by the temperature change were  $-0.32 \mu\text{mol/gFW}/^{\circ}\text{C}$  for *E. densa* ( $r = -0.985$ ,  $p < 0.01$ ),  $0.39 \mu\text{mol/gFW}/^{\circ}\text{C}$  for *M. spicatum* ( $r = 0.800$ ,  $p < 0.05$ ), and  $0.41 \mu\text{mol/gFW}/^{\circ}\text{C}$  for *C. demersum* ( $r = 0.900$ ,  $p < 0.01$ ), respectively.  $\text{H}_2\text{O}_2$  concentrations of different light intensity groups were nearly in parallel, higher with higher light intensity groups ( $p < 0.05$ ).

The increments of  $\text{H}_2\text{O}_2$  concentrations for the light exposed samples with respect to the dark-adapted ones are denoted in Figure 2. PAR intensity had significant impacts on the  $\text{H}_2\text{O}_2$  metabolism in macrophytes. For all species,  $\text{H}_2\text{O}_2$  concentrations for different light intensity groups were plotted nearly in parallel, higher with higher PAR intensity groups ( $p < 0.01$ ).

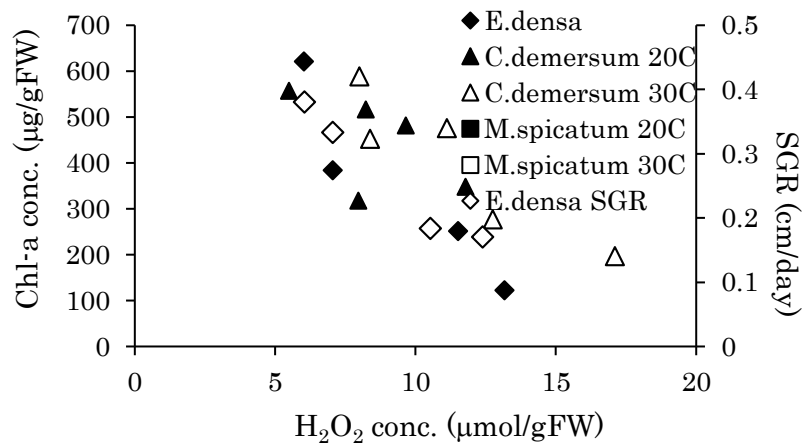


**Figure 1.** Tissue H<sub>2</sub>O<sub>2</sub> concentration vs. water temperature



**Figure 2.** Tissue H<sub>2</sub>O<sub>2</sub> concentration vs. PAR intensity (Equations are in MODELING)

These results indicate the independence of these stresses in the activity to generate H<sub>2</sub>O<sub>2</sub> at least in practical level, suggesting the total value by the sum of H<sub>2</sub>O<sub>2</sub> generated by each stress (Suzuki et al. 2014).



**Figure 3.** Chlorophyll- a concentration and SGR vs. tissue H<sub>2</sub>O<sub>2</sub> concentration

#### 4 MODELING

Figure 1 indicates that depending on species, H<sub>2</sub>O<sub>2</sub> concentration nearly linearly increases or decreases with increasing temperature. Temperature dependence of H<sub>2</sub>O<sub>2</sub> concentration is  $-0.32 \mu\text{mol/gFW}/^\circ\text{C}$  for *E. densa* ( $r = -0.985$ ,  $p < 0.01$ ),  $0.39 \mu\text{mol/gFW}/^\circ\text{C}$  for *M.spicatum* ( $r = 0.800$ ,  $p < 0.05$ ), and  $0.41 \mu\text{mol/gFW}/^\circ\text{C}$  for *C.demersum* ( $r = 0.90$ ,  $p < 0.01$ ), respectively.

From Figure 2, the following relationships were obtained for the generation rate by PAR intensity:

$$H_2O_{2\text{rad}}(\text{Temp}) = (I_0e^{-kz} - 40)^{2/3}/10 \quad \text{for } \geq 40 \mu\text{mol/m}^2/\text{s}, \quad E.densa$$

$$H_2O_{2\text{rad}}(\text{Temp}) = (I_0e^{-kz} - 40)^{5/6}/55 \quad \text{for } \geq 40 \mu\text{mol/m}^2/\text{s}, \quad M.spicatum \text{ and } C.demersum.$$

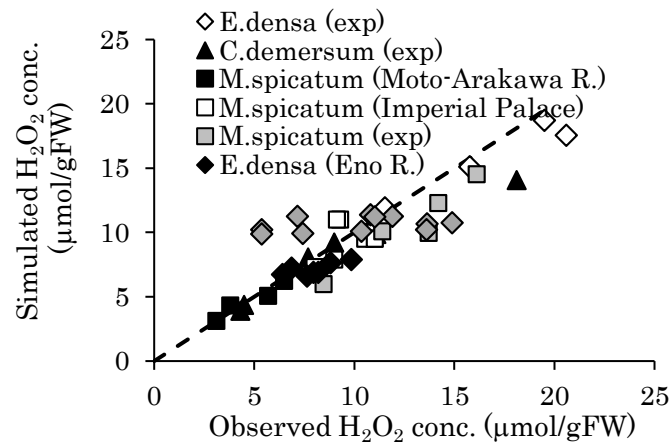
Where  $I_0$  is the PAR intensity at the water surface ( $\text{mmol/m}^2/\text{s}$ ) and  $k$  is the attenuation coefficient of light intensity in water ( $1/\text{cm}$ ).

The total H<sub>2</sub>O<sub>2</sub> concentration formed in plant tissues for a particular temperature (Temp) by the sum of H<sub>2</sub>O<sub>2</sub> generated by metabolism (H<sub>2</sub>O<sub>2met</sub>), flow velocity (H<sub>2</sub>O<sub>2vel</sub>), and solar radiation (H<sub>2</sub>O<sub>2rad</sub>). If the value is between 16 and 20  $\mu\text{mol/gFW}$ , then the growth of macrophytes deteriorates. Therefore,

$$H_2O_{2\text{tot}}(\text{Temp}) = H_2O_{2\text{rad}}(\text{Temp}) + H_2O_{2\text{met}}(\text{Temp}) < H_2O_{2\text{cr}} \quad (15-20 \mu\text{mol/gFW})$$

assuming the effects of flow velocity (H<sub>2</sub>O<sub>2vel</sub>) and other factors such as low oxygen, low nutrients are relatively small (Ellawala et al, 2011; Parveen et al. 2019; Asaeda et al. 2020; Ranawakage and Asaeda 2020).

Simulated total H<sub>2</sub>O<sub>2</sub> concentration has high agreement with the observed values (Asaeda et al. 2020, 2021).

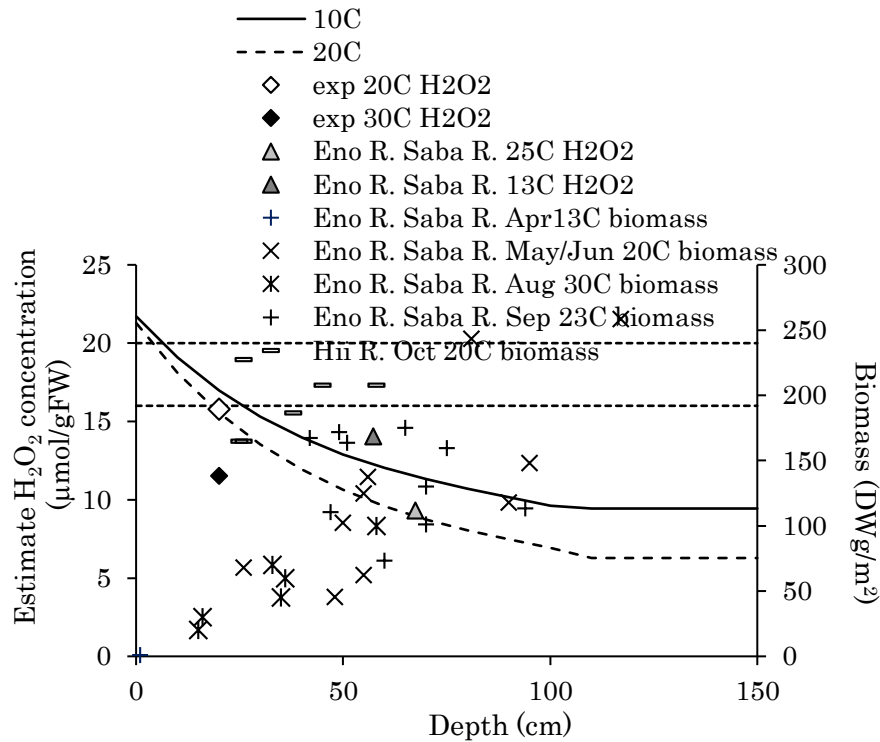


**Figure 4.** The comparison of total H<sub>2</sub>O<sub>2</sub> concentration between simulation and observation

#### 4. SIMULATED RESULTS AND DISCUSSION

Figures 5 and 6 denote the comparison between the simulated H<sub>2</sub>O<sub>2</sub> concentration with the H<sub>2</sub>O<sub>2</sub> concentration and biomass of experimental and observed results, compared with the threshold concentration, for *E.densa*, and *M.spicatum* and *C.demersum*, respectively.

Both the H<sub>2</sub>O<sub>2</sub> concentration and the existing biomass range agreed well with observed data; H<sub>2</sub>O<sub>2</sub> concentration: within 2.5  $\mu\text{mol/gFW}$ . All positive biomass range was in the range where the H<sub>2</sub>O<sub>2</sub> values were below the threshold, 16-20  $\text{mmol/gFW}$ . The H<sub>2</sub>O<sub>2</sub> concentration of *E.densa* exceeds the threshold with high PAR (Asaeda et al. 2020), thus it cannot colonize at shallow zones, while those of *M.spicatum* and *C.demersum* increases with increasing temperature, indicating the deterioration in high summer.

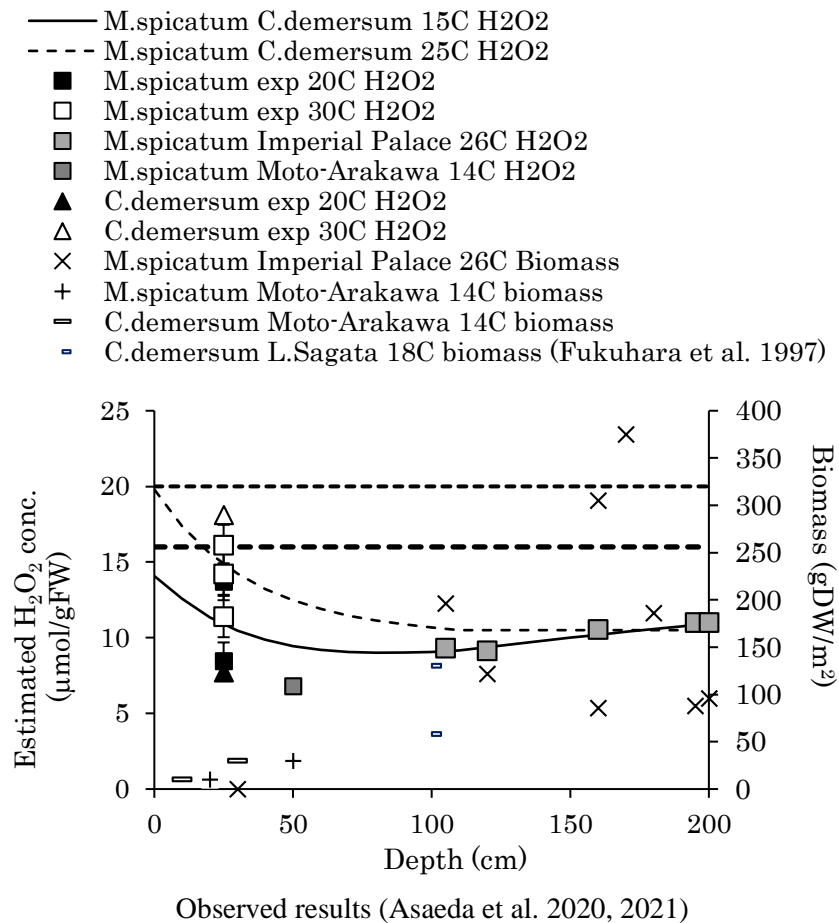


**Figure 5.** Estimated H<sub>2</sub>O<sub>2</sub> concentration vs. observed H<sub>2</sub>O<sub>2</sub> concentration and biomass for *E.densa*

Horizontal dashed lines show the threshold H<sub>2</sub>O<sub>2</sub> concentration range and observed results are (Asaeda et al. 2020, 2021).

## 5. CONCLUSIONS

Under unpreferable environmental conditions, H<sub>2</sub>O<sub>2</sub> concentrations increase in plant tissues and reflect the macrophyte condition fairly accurately. Potentially, this could be a good indicator of submerged macrophyte colonization. This approach will save time by not requiring casual observations and biomass monitoring of macrophytes in ecosystem monitoring. The experimental and field observations indicated a clear positive relationship between the level of unpreferable conditions and H<sub>2</sub>O<sub>2</sub> concentrations, regardless of abiotic factors. The total H<sub>2</sub>O<sub>2</sub> concentration is provided by the sum of H<sub>2</sub>O<sub>2</sub> generated by each environmental factor, and <16–20 μmol/gFW is required for colonization. The relationships of H<sub>2</sub>O<sub>2</sub> concentrations and the contribution of each abiotic factor were obtained for invasive species, *E. densa*, and three major Japanese native species, *M. spicatum*, and *C. demersum*. The system was applied to develop a mathematical model to simulate the colonization area of these species. The tissue H<sub>2</sub>O<sub>2</sub> concentration decreases with increasing temperature for *E.densa* and increases for other species. Therefore, native species grow intensively in spring; however, they often deteriorate in summer. For *E. densa*, on the other hand, H<sub>2</sub>O<sub>2</sub> concentration decreases with high water temperatures in summer, allowing intensive growth. High solar radiation increases the H<sub>2</sub>O<sub>2</sub> concentration, deteriorating the plant. Currently, river rehabilitation has created a deep zone in the channel, which has supported the growth and spread of *E. densa*.



**Figure 6.** Estimated H<sub>2</sub>O<sub>2</sub> concentration vs. observed H<sub>2</sub>O<sub>2</sub> concentration and biomass for *M.spicatum* and *C.demersum*

Horizontal dashed lines show the threshold H<sub>2</sub>O<sub>2</sub> concentration range and observed results are (Asaeda et al. 2020, 2021).

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### CONFLICT OF INTEREST

Author TA was employed by Hydro Technology Institute of Japan. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### REFERENCES

Asada, K. (2006). Production and scavenging of reactive oxygen species in chloroplasts and their functions. *Plant Physiol.* 141, 391. doi:10.1104/pp.106.082040

Asaeda, T., Rashid, M.H., Schoelensck, J. (2021) Tissue hydrogen peroxide concentration can explain the invasiveness of aquatic macrophytes: A modeling perspective. *Frontiers in Plant Science* 8:516301. Doi:

10.3389/fenvs.2020.516301.

- Asaeda, T., and Sanjaya, K. (2017). The effect of the shortage of gravel sediment in midstream river channels on riparian vegetation cover. *River Res. Appl.* 33, 1107–1118. doi:10.1002/rra.3166
- Asaeda, T., Sanjaya, K., and Kaneko, Y. (2017). Effects of mechanical stressors caused by mean flow and turbulence on aquatic plants with different morphologies. *Ecohydrology* 10, e1873. doi:10.1002/eco.1873
- Asaeda, T., Senavirathna, M. D. H., and Vamsi Krishna, L. (2020). Evaluation of habitat preferences of invasive macrophyte *Egeria densa* in different channel slopes using hydrogen peroxide as an indicator. *Frontiers in Plant Sciences* 11, 422. doi:10.3389/fpls.2020.00422
- Bates, A. E., Mckelvie, C. M., Sorte, C. J., Morley, S. A., Jones, N. A., Mondon, J. A., et al. (2013). Geographical range, heat tolerance and invasion success in aquatic species. *Proc. Biol. Sci.* 280, 20131958. doi:10.1098/rspb.2013.1958
- Biggs, H.J., Nikola, V.I., Gibbs, C.N., Cameron, S.M., Papadopoulos, K., Stewaart, M., Fraser, S., Vettori, D., Savio, M., O'Hara, M.T., Kucher, M., and Hicks, D., M. (2019). Flow interactions with an aquatic macrophyte: a field study using stereoscopic particle image velocimetry. *J. Ecohydraulics* 4, 113-130. Doi. Org/10.1080/24705357.2019.1606677
- Choudhury, F. K., Rivero, R. M., Blumwald, E., and Mittler, R. (2017). Reactive oxygen species, abiotic stress and stress combination. *Plant J.* 90, 856–867. doi:10.1111/tpj.13299
- Das, K., and Roychoudhury, A. (2014). Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Frontiers in Environmental Sciences* 2, 53. doi:10.3389/fenvs.2014.00053
- De Silva, H.C.C., and Asaeda, T. (2017) Effects of heat stress on growth, photosynthetic pigment, oxidative damage and competitive capacity of three submerged macrophytes. *J. Plant Interactions* 12, 228-236. doi.org/10.1080/17429145.2017.132153
- Ellawala, C. K., Asaeda, T., and Kawamura, K. (2011). The effect of flow turbulence on plant growth and several growth regulators in *Egeria densa* Planchon. *Flora* 206, 1085–1091. doi:10.1016/j.flora.2011.07.014
- Fukuhara, H., Tanaka, T., and Izumi, M. (1997). Growth and turion formation of *Ceratophyllum demersum* in a shallow lake in Japan. *Jpn. J. Limnol.* 58, 335–347.
- Satterfield, C. N., and Bonnell, A. H. (1955). Interferences in titanium sulfate method for hydrogen peroxide. *Anal. Chem.* 27, 1174–1175. doi:10.1021/ac60103a042
- Parveen, M., Miyagi, A., Kawai-Yamada, M., Rashid, M.H., and Asaeda, T. (2019) metabolic and biochemical responses of *Potamogeton anguillanus* Koides (Potamogetonaceae) to low oxygen concentrations. *J. Plant Physiol.* 232, 171-179. doi.org/10.1016/j.jplph.2018.11.023
- Queval, G., Hager, J., Gakiere, B., Noctor G., 2008. Why are literature data for H<sub>2</sub>O<sub>2</sub> contents so variable? A discussion of potential difficulties in the quantitative assay for leaf extracts. *J. Exp. Bot.* 59, 135-146.
- Sharma, P., Jha, A. B., Dubey, R. S., and Pessarakli, M. (2012). Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *J. Bot.* 26, 2012. doi:10.1155/2012/217037



- Ranawakage, V.P., and Asaeda, T. (2020). Evaluation of the physiological alterations in *Ceratophyllum demersum* L along a diurnally changing solar irradiance gradient. *J. Plant Interactions*. 15, 8-18. Doi. 10.101080/17429145.2020.1719223.
- Suzuki, N., Rivero, R. M., Shulaev, V., Blumwald, E., and Mittler, R. (2014). Abiotic and biotic stress combinations. *New Phytol*. 203, 32–43. doi:10.1111/nph.12797
- Vettori, D., Rice, S.P. (2020). Implications of environmental conditions for health status and biomechanics of freshwater macrophytes in hydraulic laboratories. *J. Ecohydraulics* 5, 71-83. doi.org/10.1080/24705357.2019.1669496
- Wellburn, A. R. (1994). The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. *J. Plant Physiol*. 144, 307–313. doi:10.1016/S0176-1617(11)81192-2